

A Set of Rearrayed BAC Clones

Spanning the Human Genome

Martin Krzywinski, Jacquie Schein, Ian Bosdet, Duane Smailus, Calum MacAulay, Wan Lam, Steven Jones, Marco Marra

Global rearray statistics are shown in Table 1

The average map size of the rearray clones is

approximates the distribution of clones in the

Figure 1 & 2. Size and band distributions showing all clones

Below are neighbour overlap statistics based on

fingerprint comparisons were carried out using a

3500

3000

2000 -

1500

1000

500

⊋ 2500 -

fingerprints in the human physical map. All

(black) and clones which failed size/bands filter (red)

REARRAY CLONE SIZE AND BANDS

2000

⊋ ¹⁵⁰⁰

s 1000

500

0 10 20 30 40 50

shared

0 10 20 30 40 50

bands (n)

neighbours (average shared 16, conserved

neighbouring rearray clones relate in terms of overlap

and size. Areas of the map

small clones (e.g. contigs

predominantly composed of

25000-25097) show up as a

QUALITY CONTROL

fingerprinted to provide a set

cluster on the size lag plot

Rearray clones will be

near 25 kbp.

Figure 5. Distribution of the number of

Figures 6 & 7 show how

shared conserved bands between

conserved

bands (n)

147 kbp and 34 bands. This very closely

100 150 200 250

size (kbp)

tolerance of 7 std mobility units.

human physical map.

3000

2 500

Z 2000

% 1500

1000

overlap (-log(Z))

scores between rearray neighbours

British Columbia Cancer Agency Vancouver, British Columbia, Canada

Genome Sciences Centre

www.bcgsc.ca info@bcysc.ca

As a subset of map clones, the

the map. On average, a

rearray represents 99.4% of

the fingerprint fragments in

canonical map clone will overlap by 75% of its size with a rearray

clone at a Sulston score of log Z

= -14.4. 1X and 2X coverage by

rearray clones accounts for 91%

of the map and is proportioned

equally - this feature increases

the effective resolution of any

hybridization or microarray

experiments.

Abstract

We have constructed a rearrayed set of BAC clones from the RPCI-11 and CalTechD libraries. The clones have been chosen from the human BAC physical map constructed at Washington University Genome Sequencing Centre. Our aim was to completely cover the entire genome with these BACs, as represented in the physical map, with a controlled degree of redundancy by using non-buried clones found in map contigs as the domain.

Our set of 29,052 size-filtered clones contains 99.6% of the fingerprint fragments found in the map and 97.5% of sequence found in the map. The rearray achieves 1X coverage for 45% of the map and 2X coverage for 46% of the map. Average coverage for any map clone by the best match in the rearray is 75% (108 kbp) with an average Sulston score of 10⁻¹⁴.

We anticipate this resource will have several uses, including provision of a genome-ordered set of probes for fluorescent in-situ hybridization and provision of probes for microarray-based BAC-Comparative Genomic Hybridization (BAC-CGH) experiments. The rearray clone set will be fingerprinted to verify clone identity and to provide a high-quality map of the resource. Future plans include creating similar lists for mouse, rat, poplar and bovine genomes.

Clone Set Creation Algorithm

The human BAC physical map is a curated and sequence-validated resource, well suited for selection of a minimal set of clones spanning the genome. Using the Nov 2001 version of the map, our approach involves walking along each contig and selecting clones in the following fashion

0. Repeat-masked BAC clone end sequences are searched by BLAST against repeatmasked human sequence (Aug 2001) to determine location of the clone, where possible. Hits with $-\log(\text{Expect}) > 20$ or those with $-\log(\text{Expect}) > 50$ and a match fraction of less than 80% are considered weak and discarded. For any clone, if BAC ends are closer than 2 kbp or further than 5 Mbp apart, they are discarded.

OUTER LOOP: foreach contig in map

1. All "non-buried" clones (canonical) in a contig are ordered by their left end positions (secondary sort by right end). Clones from RPCI-11 and CaltechD are marked as available and are eligible for the rearray. All other clones are considered unavailable. Clones that are buried or singletons (those not assigned to a contig) are not considered and are disregarded.

2. All admissible BAC end hits are used to determine the consensus chromosome for the map contig. Any clones with strong hits to other chromsomes are marked as inadmissable for the rearray (failed consensus). Out of the remaining clones, any clones farther than the length of the contig from the statistical mode BAC end position are also inadmissable. Hit polarity is used to determine left/right orientation of clones with only one admissable BAC end hit if a clone with both ends hits to the same sequence contig. Any clones whose single end cannot be categorized as left/right is marked as a middle hit and not used for overlap determination.

3. All admissible clones outside of the range of 100-200 kbp or 20-50 bands are marked and selected towards the rearray set only in places where a lack of coverage would otherwise occur.

Maximized quantities

Minimized quantities

High degree of overlap

4. The left-most clone is used as a starting point for a walk. To maximize coverage, the



Rearray Statistics

29,052 clones

95% RPCI-11 5% Caltech D1/D2

20% sequence clones (full X)

5% failed size/band filter **52%** have BAC end hits **42%** both ends **10%** left only 12% right only 36% unclassified

19% overlap by sequence with next pick

4.3 Gbp total size 2.9 Gbp unique band size

Table 1. Global rearray statistics.



50 100 150 200 overlap (kb) Figure 4. Distribution Sulston Figure 3. Distribution of overlap

between rearray neighbours, as determined by the size of shared bands (average 71 kbp).





(average 6.4).



Figure 8. Distribution of the number of Figure 9. Distribution of left end same contig hits among top 10 between map and rearray clones (black). The rank of the highest scoring same contig nit is shown in orange.





Rearray Clone Overlap (%) Figure 11. Distribution of the fractional overlap (sequence) between rearray neighbours (average 75.5, 95% clones >55% overlap, 90% clones >60% overlap)

Rearray Clone Overlap (kbp) Figure 12. Distribution of the absolute overlap (sequence) betwee rearray neighbours (average 108 kbp, 95% clones > 74 kbp, 90% clones > 81 kbp).

About 110 RPCI-11 canonical map BACs associated with subtelomeric regions were matched to the rearray. For about half of these clones, some telomere distance information was available. Figure 13 shows the representation of these clones in the rearray.





Determination of Sequence Coverage

Sequence coverage was evaluated using in-silico restriction digests from human sequence, counting the number of restriction fragments represented in the rearray. All possible non-overlapping 100 kb fragments without sequence gaps were created from sequence (17,000, approximately 56% of the genome) and digested in-silico with HindIII. Junction fragments and fragments smaller than 600 bp were removed and multiplets within 7 std mob units were collapsed to the largest fragment. This sanitization was done

Best Match Score (-log(Z)) Figure 10. Distribution of the Sulston score. Z. for the same best contig hit (average 14.4). The cumulative listribution is shown in green (95% clones >7.5, 90% clones >8.8)







walk always starts at the first available clone, regardless of size, band or BAC end information. The following loop is iterated until the end of the contig is reached.

INNER LOOP: foreach clone in contig

5. A forward-looking local neighbourhood of
clones is selected. This neighbourhood extends 20 cb
map units past the current clone. For each clone in
the neighbourhood, we determine the number of
conserved bands (bands shared by the clones and all
clones in between) and sequence overlap by using
any available BAC end information.

6. From the neighbourhood clones we pick a single clone as the walk destination. Clones which pass the size/bands filter are always considered first. The evaluation proceeds in this order

i. right-most clone with sequence overlap is chosen

ii. right-most clone with admissible BAC ends is chosen. Clones with hits for both BAC ends must be at least 5 CB map units to the right. Clones with single BAC end hits must be at least 10 CB map units to the right.

iii. right-most clone with fewest conserved bands >4 is chosen

iv. if no such clones can be found, the next available clone is chosen, regardless of any size, band or BAC end information.

7. If the optimal clone, as determined solely by map position, fails size, band or BAC end position filters, then the next clone is chosen if overlap is not sacrificed. The algorithm is designed to minimize the number of sub-quality clones in the rearray, but choses them if they are the only alternative.



For each map clone's top 10 hits, the highest ranking hit on the same contig was used for Figures 9-12. Only 0.6% of map clones did not show same contig hits in the top 10 list. These are (i) typically shorter clones (average 126 kbp) whose coverage is split between two rearray clones, or (ii) unavailable clones which do not overlap available clones.



to mimic as closely as possible the sanitization state of clones in the fingerprint map. As a control, the same comparison was done for all canonical map clones, as well as for all available map clones.



Figure 17. Distribution of resentation difference and number o unmatched bands between the map and rearray clone set. Negative differences can be explained by the fact that the local neighbourhood for the best match in the map and in the rearray may be



External Resources

Clone libraries RPCI-11: Osoegawa et al. (2001) Genome Res 11(3):483-496), CalTechD: Knight and Lese et al. (2000) Am J Hum Genet, 67(2):320-332

Fingerprinting Marra M etal. (1997) Genome Res 7:1072-84

rearray cannot represent more sequence than the map, being a

and removes dependance on the algorithm parameters. With

this method, we find that the rearray achieves 97.5%

sequence coverage, when compared to the map.

subset of the map, normalizing by the control result is justifiable

BAC physical map Washington University Genome Sequencing Centre [www.genome.wustl.edu]

Golden path generation: International Human Genome Sequencing Consortium news/pr/jun2000/nhgri-26.html], assembly: GigAssembler [genome.ucsc.edu/goldenPath/algo.html] by Jim Kent

BAC End database: TIGR [www.tigr.org]

Human Telomere Mapping and Sequencing Project Riethman H et al. [www.wistar.upenn.edu/Riethman]

Genome Sciences Centre Team

Bioinformatics Jacquie Schein, Chris Fjell, Steven Jones, Marco Marra Laboratory Ian Bosdet, Duane Smailus, Carrie Mathewson, Natasja Wye Associates Wan Lam, Calum MacAuley, Adrian Ishkanian

Funding

Genome Canada Cancer Genomics Project: Victor Ling, Connie Eaves, Marco Marra

ounding Director

Dr. Michael Smith